

MALDI-MS Screening of Microbial Colonies with Isomer Resolution to Select Fatty Acid Desaturase Variants

Background/Objective

Controlling double bond locations in oleochemicals can be a key to producing many value-added products in microbes. However, measuring double-bond locations for improving production is not currently amenable to high-throughput assays due to isobaric masses of the double-bond isomers. We developed a mass spectrometry (MS) assay to determine the positions of double bonds on membrane lipids.

Approach

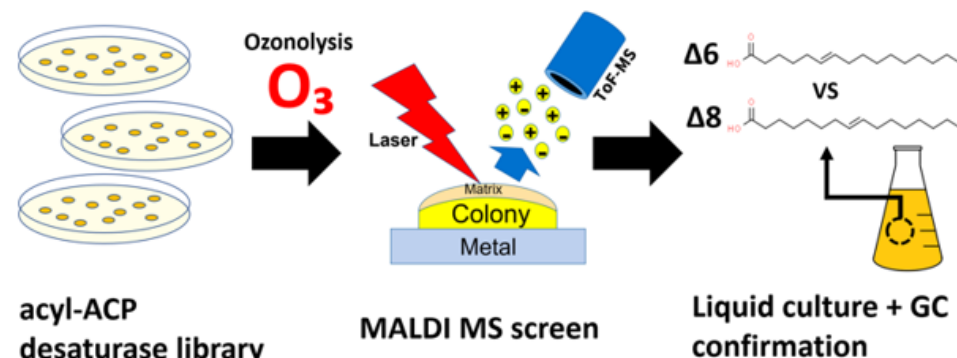
- Agar colonies of *E.coli* expressing a randomly mutagenized library of a C16:1 $\Delta 6$ *Thunbergia alata* desaturase gene were treated with ozone.
- Matrix-assisted laser desorption ionization (MALDI) MS was used to measure ozonolysis products of unsaturated fatty acids in each colony for 5 seconds.
- Colonies showing increased C16:1 $\Delta 8$, indicating an increase in activity, were validated by liquid culture and gas chromatography (GC).

Results

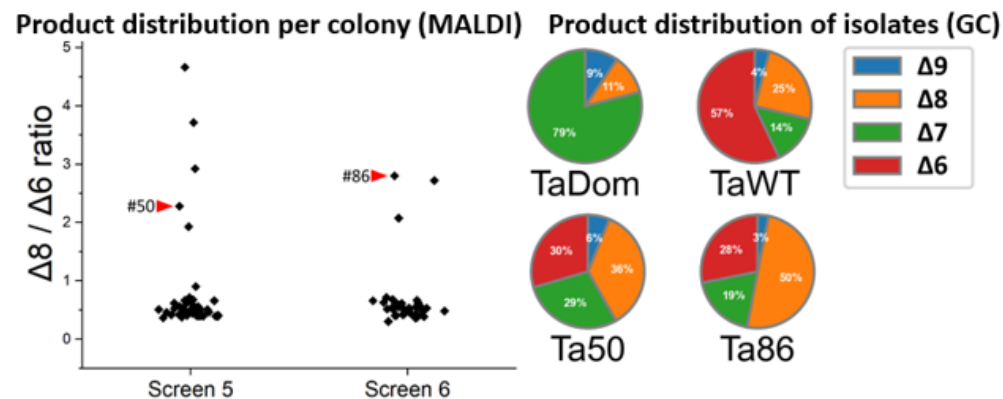
GC analysis for the liquid cultures of the isolated colonies showed dramatically increased ratio of $\Delta 8$ double bonds in the C16:1 species, validating the screen.

Significance/Impacts

This provides a simple and economical high-throughput screening method for improved enzyme activity and metabolic circuits toward producing desired unsaturated fatty acids.



MALDI screen isolated regioselectivity variants



Overall workflow of MALDI-MS screening of the $\Delta 6$ desaturase random library and validation of $\Delta 8$ -active variants by GC-MS.